DNAMAN Sequence Analysis Software

Oligo Sequences Analysis and PCR Primers

BIOINFORMATICS PLATFORM

Oligo Sequences and PCR Primers

Oligo Sequence Analysis
 Oligo Secondary Structure
 PCR Primers

Enter Oligo Sequence



	i - Tati a	
Primer	1	×
Enter primer sequence	•	
CCACTTTCACAGGCACAGGAGCA		
ОК	Cance	1

Oligo sequence can be entered:

- 1. Directly from Enter box
- 2. From Oligo database

Enter Oligo Sequence



	i - Tati a	
Primer	1	×
Enter primer sequence	•	
CCACTTTCACAGGCACAGGAGCA		
ОК	Cance	1

Oligo sequence can be entered:

- 1. Directly from Enter box
- 2. From Oligo database

Oligo Properties



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- 1. Select Oligo Properties tool
- Properties shown in a dialog box.
 Oligo sequence can be modified.
- Press Report button to show properties in Text window

Melting Temperature		
Oligo seque	ince	
CCACTTT	CACAGGCACAGGAGCA	2
L.	ength 23 GC% 56.5	5 MW(kD) 7.04
	Melting Temperature (°C):	
Thermo	64.2 Hybrid 67.0	GC+AT 72.0
1 D]	VA] (nM) 50	
[N;	a+](mM) 200	Formamide(%) 0 Mismatch(bp) 0
	Show Tm Repo	rt <mark>3</mark> <u>C</u> ancel

Secondary Structure



PCR Primers



PCR Primers Step 1: Primer Filters

Step 1 of 3: Primer filtrat	tion
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Primer locations on target seque	ence	4	Primer properties
Product size (bp) from Sense primer from Antisense primer from	400 to 800 1 to 886 1 to 886		▼ Shortest primers ▼ G/C at 3' end Length (base) 18 to 22 Tm (°C) 56 to 62 GC (%) 45 to 55
Reject primers 3' Dimers (bp) > PloyN (base) > Primer-Primer: Continuous(bp)>	3 3 3	Hairpin Stem (bp) > 3 3' Unique (base) < 6 All matches(%)> 5(Concentrations for Tm calculation Primer (nM) 50 Salt (mM) 200
Product for hybridyzation	Product Tm(°C) range from Product GC content (%) from	70 to 90 40 to 70	Hybridization [Salt] (mM) 200 Exclude product with polyN (bp) > 8

- 1. Select primer locations on template DNA
- 2. Set primer properties with component concentrations
- 3. Set Rejection metrics to filter low quality primers
- 4. Set PCR product properties if the product will be used for hybridyzation

PCR Primers Step 2: Primer Filters

Step 2 of 3: Refinement and pair selection

13 Sense primers	Export List	8	Antisense primers
29 TTACTGCCAAGGACATTCTGG 56.1°C 32 CTGCCAAGGACATTCTGGAC 56.4°C 33 TGCCAAGGACATTCTGGAC 57.8°C 94 TTCTATGTGGCAAGACGACG 56.9°C 189 TGATAGCAAAGCCAAGACGAAG 57.6°C 193 AGCCAAGACGAAGACGAAG 56.1°C 194 AAAGCCAAGACGAAGACGAAG 56.1°C 198 AGCCAAGACGAAGACGAAGA 57.4°C 320 CCTAAACCAGATGACGGCTG 56.2°C 323 AAACCAGATGACGGCTGC 56.9°C 323 AAACCAGATGACGGCTGCTA 58.9°C 476 ACTTTCAACGGATTCCAGAGG 56.2°C 478 TTTCAACGGATTCCAGAGGC 56.0°C	497 526 533 536 540 874 878 880	GCCTCTGGAATCCGTTGAAAG GCACAGGAGCATCGCATAATG TTCACAGGCACAGGAGCATC ACTTTCACAGGCACAGGAGG TTCCACTTTCACAGGCACAG CCCATTCTGCCATACACAAGC TTAGCCCATTCTGCCATACAC GCTTAGCCCATTCTGCCATAC	57.5°C 58.3°C 58.2°C 58.4°C 56.2°C 57.8°C 56.3°C 57.3°C
Reject primer pairs Primer Tm difference (*C) > Mispriming analysis on target sequence Out off% >=	1 •	No restriction analysis	 Keep primers with restriction site Keep primers without restriction site

Sense and anti-sense primers are selected. Set parameters to select pairs. 1. Set parameters to reject pairs with different Tm or mispriming target DNA

- 2. Set restriction site requirement to allow or reject primers
- 3. All primers can be exported to a Text window for records

PCR Primers Step 3: Primer Pairs

Step 3 of 3: Final



- 1. Select a pair of primers visualize the properties
- 2. Primer pair locations shown in the target DNA diagram
- 3. Product length and primer information
- 4. Primer quality can be sorted in Position/Product Size/Tm

PCR Primers Final: Primer Pair List

